In vitro Antiplasmodial and Haemolytic Activities of Trema orientalis, Cnestis ferruginea and Dialium dinklagei Used to Treat Malaria in Côte d’Ivoire

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Authors’ contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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ABSTRACT

Malaria constitutes one of the biggest health problems in tropical Africa due to the resistance of human malaria parasites to anti-malarial compounds. Research focused on plants used in traditional medicine to treat malaria is still a viable alternative for the creation of novel anti-malarial drugs. This study evaluated extracts from three medicinal plants, Trema orientalis, Cnestis ferruginea and Dialium dinklagei, used in traditional medicine in Côte d’Ivoire, for in vitro antiplasmodial activities. SYBR GREEN fluorescence method was used to evaluate the in vitro...
inhibitory activity of the extracts, chloroquine, artemunate and quinine against *Plasmodium falciparum* field isolates and two laboratory strains of *Plasmodium falciparum*: the chloroquine sensitive 3D7 and the chloroquine resistant Dd2. In comparison to plant extracts, chloroquine, quinine, and artemunate were chosen as reference antimalarials. In addition, the haemolytic activity of extracts showing good antiplasmodial activity was evaluated. The IC₅₀ and the corresponding correlation coefficients were determined graphically, using *In vitro* Analysis and Reporting Tool (IVART) software of WWARN (Worldwide Antimalarial Resistance Network). Results showed that no plant was active with the hexanolic extract. *Trema orientalis* had moderate activity with the methanolic extract with activities ranging from 14.46µg/mL to 28.32µg/mL. *Cnestis ferruginea* was active with the decoction extracts with activities ranging from 11.78µg/mL to 13.94µg/mL. *Dialium dinklagei* was active with both methanolic and aqueous extracts ranging from 12.80µg/mL to 21.67µg/mL. There was less than 1% hemolysis at the concentration of 200 µg/mL of plant extracts. These results validate the reported traditional use of *Trema orientalis*, *Cnestis ferruginea* and *Dialium dinklagei* for malaria treatment in Côte d’Ivoire.

Keywords: Antiplasmodial; haemolytic; *In vitro*; malaria; *Plasmodium falciparum*.

**1 INTRODUCTION**

In 2019, approximately 229 million cases with 409,000 deaths attributable to malaria have been estimated [1]. “These alarming statistics make malaria the world’s leading endemic. Chemotherapy treatment of malaria has evolved over the past decade due to the spread of multirresistant strains of *Plasmodium falciparum*. The World Health Organization (WHO) is currently promoting artemisinin-based combination therapy (ACT) as the reference drug for the management of uncomplicated falciparum malaria to reduce the risk of resistance” [2,3].

“Appearance and rapid extension of *Plasmodium falciparum* resistance to the common antimalarial agents such as sulfadoxine-pyrimethamine, chloroquine and recently to the derivatives of artemisinin makes urgent the discovery of new antimalarial compounds” [2,4,5].

“Plants have been and remain a good source of pharmacologically active compounds, including antimalarial agents, as shown by quinine, isolated from *Cinchona* sp. and artemisinin extracted from *Artemisia annua*. In addition, many phytochemical compounds with antimalarial activity were isolated from plants” [6,7].

The majority of the population still using traditional medicines to cure diseases like malaria despite the availability of modern treatments. The high cost of treatment and frequently cultural considerations are the most frequently mentioned causes. “In Côte d’Ivoire, a large number of plant species have been identified as antimalarial medicinal plants. Pure products have been isolated from some of these plants including those with antimalarial activities comparable to or more active than chloroquine on susceptible and resistant strains of *P. falciparum*” [8,9]. It is therefore imperative to pursue antimalarial drug development with these highly active products through to drug production. This led us to study *Cnestis ferruginea Vahl*, *Dialium dinklagei* and *Trema orientalis*.

*Dialium dinklagei* belongs to the *Fabaceae* family. It is a plant found in forest areas from Guinea to Congo. This plant is used in Côte d’Ivoire to treat malaria. Very little work has been done on this plant, but we note that Bouquet and Debray have described the presence of tannins in the leaves of this plant [10].

*Cnestis ferruginea* Vahl ex DC is a perennial shrub or tree belonging to the family *Conaraceae*. *Cnestis ferruginea* has a wide distribution in West Africa, particularly in Gambia, Ghana, Guinea-Bissau, Côte d’Ivoire, Liberia, Nigeria, Sierra Leone [11], Benin [12], Niger and Gabon, especially in semi-deciduous forests.

The leaves are highly effective vermifuges against ascaris, claim Bouquet and Debray. The leaves also have purgative qualities and are used to cure asthma and scabies. Another therapeutic indication that the traditional pharmacopoeia recognizes to this plant is its use for the treatment of the ocular affections. Several other uses were described by authors [10].

*C. ferruginea* is well known in phyotherapy and certain literatures lend it diverse therapeutic uses such as the treatment of conjunctivitis, bronchitis, tuberculosis, migraines, sinusitis and oral...
infection. It is also used for the treatment of dysentery, syphilis, gonorrhea, cough, dysmenorrhea, ovarian and aphrodisiac disorders, abortion and constipation [13]. The leaves can be used as a laxative and against fever [14]. The roots and fruits have been considered as a remedy for snake bite. The decoction of the bark can be used to treat gum infections [11].

The fruits are also used in ocular pathologies such as conjunctivitis and against several other diseases like bronchitis or tuberculosis [14,15]. Several works have shown the anti-inflammatory and analgesic activities [16,17] and the anti-depressive and anxiolytic activities [18] of *Cnestis ferruginea*.

*Trema orientalis* is an arborescent species belonging to the family *Ulmaceae*. In Africa, its geographical area extends from Senegal to Somalia, through the entire region of Central Africa.

*Trema orientalis* is generally used to treat jaundice, broncho-pulmonary affections, fever, rheumatic pains and malaria. Administered orally, the plant is reported to have a purgative and diuretic action [10].

Bouquet and Debray noted the absence of alkaloids [10]. As for Bekro et al, they noted an absence of gall tannin, quinone and saponosides [19]. Kerharo and Adam reported that the leaves exert diuretic effects due to flavonoids and vermifuge effects due to polyterpenes [15]. Dimo et al. also reported that “*Trema orientalis* has anti-diabetic properties (hypoglycemic activity) and could be beneficial for diabetics and other cardiovascular diseases” [20]. Dijoux-Franca et al. showed “the presence of dihydrophenanthrene and phenylidihydroisocoumarin in *Trema orientalis*” [21].

2 MATERIAL AND METHODS

2.1 Plant Material

The plant material consists of the leaves of *Dialium dinklagei*, *Cnestis ferruginea* and *Trema orientalis*. The leaves of these plants were collected since March 2013 in the District of Abidjan.

The timing of the plant harvest was the morning at 9 AM.

2.2 Biological Material

Biological samples are constituted of group O blood samples with a positive rhesus (Rh+) for an inoculum dilution with clinical and reference strains of *Plasmodium falciparum*.

Two ATCC reference strains were used: 3D7 chloroquine-sensitive was provided by Biochemistry and Molecular Department University of Legon, Ghana and Dd2 chloroquine-resistant was provided by MRA-156, lot N°58319486, MR4ATCC®Manassas, Virginia, USA.

2.3 Methods

2.3.1 Ethnobotanical survey, bibliographic research and selection of studied plants

Popular pharmacopoeia methods were used to establish a list of plants used to treat malaria. Popular pharmacopoeia [22] consists of direct, collective or individual interviews with populations on the plants used in routine care.

Investigations were carried out in November 2012 in Abidjan and Bondoukou (city in the east of Côte d’Ivoire) by ethnobotanical approaches with 27 actors of traditional medicine [22]. Ethnobotanical data (local name, method of preparation, traditional use, combination of plants, indications, dosage, contraindications and side effects) are obtained through conversations with traditional healers.

Samples collected were identified at Centre National de Floristique (National Floristry Center) of Félix Houphouët-Boigny University, Abidjan, (Côte d’Ivoire) by Professeur Aké-Assi Laurent.

2.4 Extracts Preparation

The leaves were dried out of the sun for one week at room temperature before being reduced to fine powder using a mechanical grinder (Retsch M6951). From the powder obtained, the various crude extracts were prepared. Decoction of each plant was made as close as possible to the traditional healer’s formula. Then, 3 successive extractions by solvents of increasing polarity (hexan, methanol and water), have been done according to the protocols made by Zirihi et al. et Bekro et al. [19,23] (Figs. 1 and 2).
Fig. 1. Diagram showing leaves extracts by decoction

- 20 g of plant powder in 1 L of distilled water
- Ebullition for 30 minutes then filtration by hydrophilic cotton and Whatman paper
- Marc
- Filtrat 1
- Addition of 1 L of water distilled boiling for 30 minutes then filtration on cotton hydrophilic and whatman paper
- Filtrat 2
- Evaporation at 55°C for 48 Hours
- Decoction extract

Fig. 2. Diagram showing extraction of leaves using hexane, methanol and water [19,23]
2.5 Field Isolates Collection

Blood samples were collected at the health center of Wassakara (Abidjan Côte d’Ivoire) by venipuncture in heparinized tubes from patients older than 18 years with uncomplicated *P. falciparum* malaria after informed consent. The samples were then transferred at 4°C to the Swiss Center for Scientific Research for *in vitro* test.

2.6 *In vitro* Antiplasmodial Assay

“Antiplasmodial activity was analyzed with SYBR Green method. The assays were carried out on 96-well plates filled with a infected red blood cells (IRBCs) in the following proportions of parasitaemia <0.3% and hematocrit 5%. The *in vitro* *P. falciparum* continuous culture used in our assays is derived from that developed” by Trager et Jensen [24]. “Inhibition of parasite growth was measured using the SYBR Green method” [25–29].

“The reading was done with the Spectra Max GEMINI XPS spectrofluorometer (Molecular Devices) at 535 nm after excitation at 485 nm. The IC50 are determined graphically, using *In Vitro* Analysis and Reporting Tool (IVART) software of WWARN” [26, 29]. The extracts are then qualified as follows:

- $\text{IC}_{50} > 50 \text{ µM}$, the compound is considered inactive.
- $11 < \text{IC}_{50} < 50 \text{ µM}$, the compound is not very active.
- $2 < \text{IC}_{50} < 11 \text{ µM}$, the compound is active.
- $\text{IC}_{50} < 1 \text{ µM}$, the compound is highly active and can be considered as a “lead compound” [30,31].

2.7 *In vitro* Hemolysis Assay

A stock solution of the samples was prepared in an appropriate solvent at concentrations of 100 µg/mL and 50 µg/mL, taking into account that the solvent volume must not be greater than 1% in the final solution.

To perform hemolysis assay, 10 µL of stock solution was placed in an Eppendorf microtube and mixed with 190 µL of RBC (10%) as controls. The negative control comprised 10 µL of PBS + 190 µL of 10% RBC and the positive control was prepared with 10 µL of 20% Triton X-100 + 190 µL of 10% RBC. Tubes were centrifuged for 5 minutes at 2200 rpm and 150 µL of supernatants were placed in a 96-well plate. The absorbance was read at 550 nm with a plate reader (Multiskan FC, Thermo Scientific).

The following formula was used to calculate the percentage of hemolysis: 

$$\% \text{ hemolysis} = \left(\frac{(\text{Abs sample} - \text{Abs negative control})}{(\text{Abs positive control} - \text{Abs negative control})}\right) \times 100$$

Abs= absorbance at 550 nm

3. RESULTS

3.1 Antiplasmodial Activity

The results showed that no plant is active with the hexanolic extract. *Trema orientalis* had moderate activity with the methanolic extract with activities ranging from 14.46 µg/mL to 28.32 µg/mL. *Cnestis ferruginea* was active with the decoction extracts with activities ranging from 11.78 µg/mL to 13.94 µg/mL. *Dialium dinklagei* was active with both methanolic and aqueous extracts.

In summary, decoction extract of *Cnestis ferruginea* and *Dialium dinklagei* showed good activity against fields and references parasites. The methanolic extract of *Trema orientalis* had moderate antiplasmodial activity. The four field isolates tested were CQ sensible. Quinine and Artesunate showed good activity against field isolates (Table 1).

3.2 Haemolytic Activity

No extract was found to exhibit significant red blood cells lysis activity with a percentage of haemolysis <1% for all tested extracts (conc = 100 µg/mL and 200 µg/mL). This indicates that anti-plasmodial activity is not correlated with haemolysis of red blood cells but with a real action against the parasite.

4. DISCUSSION

The study of the anti-plasmodial activity of a plant requires many analytical tools. In the current study, the effectiveness of a plant is demonstrated by the *in vitro* activity of one of its extracts against *P. falciparum*. This study investigated the *in vitro* activity of extracts of three plants on *Plasmodium falciparum* reference strains (Dd2 and 3D7) and clinical strains (W536, W539, W552, ANK02). This work is justified by the emergence and spread of *P. falciparum* resistance to current antimalarial drugs. Screening of plants used in traditional medicine for antiplasmodial activity is one way to discover new effective agents” [9,32].
Table 1. Antiplasmodial activity of crude extracts

<table>
<thead>
<tr>
<th>Plants</th>
<th>Extracts</th>
<th>Extraction yield (%)</th>
<th>W536</th>
<th>W539</th>
<th>W552</th>
<th>ANK02</th>
<th>3D7</th>
<th>Dd2</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Dialium dinklagei</strong></td>
<td>Déc</td>
<td>18.5</td>
<td>13.19±3.17</td>
<td>13.74±4.12</td>
<td>12.8±2.4</td>
<td>12.9±3.01</td>
<td>14.35±0.79</td>
<td>13.82±2.7</td>
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<tr>
<td></td>
<td>Hex</td>
<td>1</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
</tr>
<tr>
<td></td>
<td>Met</td>
<td>4.5</td>
<td>17.66±2.65</td>
<td>15.76±5.63</td>
<td>19.15±3.27</td>
<td>15.29±3.76</td>
<td>14.97±2.59</td>
<td>15.11±2.26</td>
</tr>
<tr>
<td></td>
<td>Aq</td>
<td>4</td>
<td>18.34±3.01</td>
<td>15.43±2.24</td>
<td>17.33±4.21</td>
<td>21.67±3.29</td>
<td>19.54±1.58</td>
<td>17.47±1.26</td>
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<tr>
<td><strong>Cnestis ferruginea</strong></td>
<td>Déc</td>
<td>17.95</td>
<td>12.85±2.29</td>
<td>13.1±3.19</td>
<td>11.96±1.62</td>
<td>13.94±3.23</td>
<td>11.78±2.21</td>
<td>11.85±1.43</td>
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<tr>
<td></td>
<td>Hex</td>
<td>1</td>
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<tr>
<td></td>
<td>Met</td>
<td>5.4</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
</tr>
<tr>
<td></td>
<td>Aq</td>
<td>5.8</td>
<td>13.68±2.76</td>
<td>13.74±2.72</td>
<td>12.35±1.44</td>
<td>13.56±3.1</td>
<td>12.15±1.91</td>
<td>12.35±4.21</td>
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<tr>
<td><strong>Trema orientalis</strong></td>
<td>Déc</td>
<td>20</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
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</tr>
<tr>
<td></td>
<td>Hex</td>
<td>1.2</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;50</td>
</tr>
<tr>
<td></td>
<td>Met</td>
<td>7.2</td>
<td>14.46±2.55</td>
<td>22.54±5.67</td>
<td>19.65±3.29</td>
<td>28.32±2.34</td>
<td>19.57±1.28</td>
<td>22.65±3.39</td>
</tr>
<tr>
<td></td>
<td>Aq</td>
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<td>&gt;50</td>
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<td>&gt;50</td>
<td>&gt;50</td>
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</tr>
</tbody>
</table>

Chloroquine (nM) (0.01µg/mL) 33.01±0.92 42.71±1.32 37.31±3.24 35.38±4.92 51.07±2.23 116.71±5.11
Quinine (nM) (0.01µg/mL) 5.76±0.95 23.87±1.12 44.37±2.15
Artesunate (nM) (0.001µg/mL) 3.43±0.49 2.22±0.16 6.31±3.01

Dec : Décotion ; Hex : Hexanique ; Met : Méthanolique ; Aq : Aqueux
In vitro inhibitory activity of aqueous, methanolic and hexanic extracts of Dialium dinklagei, Cnestis ferruginea and Trema orientalis leaves on chloroquine sensitive and chloroquine resistant laboratory strains and field isolates P. falciparum were tested.

The determination of antimalarial activity on reference strains of P. falciparum also raises the problem of the validity of in vitro tests. Indeed, these different strains used are adapted to laboratory culture and may not match the reality of the environment. This is why it appeared useful to evaluate the activity of the extracts on isolates from malaria patients in endemic areas. In brief, our results indicate that decoction extract of Cnestis ferruginea and Dialium dinklagei showed good activity against fields and references parasites. The methanolic extract of Trema orientalis had moderate antiplasmodial activity.

The results showed that no plant is active with the hexanic extract.

“Cnestis ferruginea was active with the decoction extracts with activities ranging from 11.78µg/mL to 13.94µg/mL. Cnestis ferruginea has not yet been tested for its antimalarial activity, however it is well known for antibacterial property, and has been used traditionally to treat several infectious diseases” [33]. “Aqueous extracts of C. ferruginea leaves showed antimicrobial activity due to the presence of hydroquinone and caffeic acid methyl ester” [34]. The antiplasmodial activity observed in our study could be due to the presence of these chemical compounds.

“A review found that many Connaraceae species with associated pharmacological potential have traditional medicinal use in tropical areas worldwide. The traditional uses reported in the different ethnomedical studies include a wide range of therapeutic functions, which can guide research for actions that have not been confirmed yet by the scientific community” [35].

Trema orientalis had moderate activity with the methanolic extract with activities ranging from 14.46µg/mL to 28.32µg/mL. However, no activity was observed with hexanic and aqueous extracts. “Very little work has been done in vitro, however several studies have been done on the in vivo antiplasmodial activity. In fact, the other study indicates that the aqueous leaf and bark extracts of T. orientalis have good antiplasmodial activity, with varying degree and/or differential effect on the measured parameters” [35–37]. “According to these results, the acetone leaf extract of T. orientalis has antiplasmodial activities and dosage of 800 mg/kg/day is the most effective dose” [38–40]. “In the prophylactic experiment, dichloromethane, methanol fraction and extract showed significant chemopreventive effects against P. berghei invasion of the red blood cells when compared with both Sulfadoxine-Pyrimethamine and untreated controls” [41]. In addition, this study provides evidence of the claims by the traditional healers of the efficacy of aqueous extracts of T. orientalis.

Dialium dinklagei was active with both methanolic and aqueous extracts with activities ranging from 12.80µg/mL to 21.67µg/mL. According to Akoué et al, the abundance of bioactive compounds would explain the therapeutic effects observed and the use of Dialium dinklagei in traditional medicine. Abundance in secondary metabolites of Dialium dinklagei may justify their use in ethnotherapy by populations [42].

All plants species exhibited in vitro antiplasmodial activity against reference strain of Plasmodium falciparum. Some activities are less than 15 µg/mL and are considered promising according to Bero et al [30,43]. The four field isolates tested were Chloroquine sensible. Since 2003, chloroquine has been excluded from recommended regimens for treating malaria. It appeared that this chemical started acting now on P. falciparum clinical isolates. However, this reversion of chloroquine resistance should be confirmed by future In vitro and In vivo as well as by molecular studies. Quinine and Artesunate showed good activity against field isolates.

“Haemolytic activity represents a useful starting point as it provides the primary information on the interaction between molecules and biological entities at cellular level. Haemolytic activity of any compounds is an indicator of general cytotoxicity towards normal healthy cell. The results obtained in this study indicated the absence of hemolytic activity. This indicates that anti-plasmodial activity was not due to haemolysis of red blood cells but with a real effect of the extracts against the parasite. Therefore, we can conclude that the results obtained during the antiplasmodial activity are not influenced by this weak haemolytic action” [31].
5. CONCLUSION

Our experimental approach allowed to select extracts with good antiplasmodial activities and to validate their use in the traditional Ivorian pharmacopoeia for the treatment of malaria. Overall, the in vitro activities of these plant extracts have proven their use as alternative traditional remedies for malaria. Several herbs were used by traditional healers to treat malaria symptoms, however, these plants lacked antiplasmodial action in vitro. It should be emphasised that only the asexual erythrocytic stage of Plasmodium falciparum was evaluated for antiplasmodial action, and the extracts that proved inert may still inhibit later stages of the parasite. On the other hand, some plants lacking in vitro antiplasmodial action might elicit a strong immunological response. Additionally, the extracts' tests for hemolytic activity, which could conflict with the antimalarial action, revealed no hemolytic activity. Following the elaboration of some components of this research and the completion of clinical trials, this work might serve as the basis for the creation of conventionally enhanced medications for malaria treatment.

CONSENT AND ETHICAL APPROVAL

The study was conducted in accordance with the local laws and regulations, and International Conference on Harmonization - Good Clinical Practice (ICH-GCP). The protocol was reviewed and approved by the National Ethical Committee for Research (03-2013 /MSLS/CNER-P). Written informed consent was obtained from participants for blood collection and from traditional healers. In case of an illiterate participant, his/her thumb impression and signature of an independent witness were sought.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

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